ORIGINAL PAPER

Two Highly Sensitive and Selective Colorimetric "Off-On" Rhodamine-Based Fluorescent Chemosensor for Hg(II) in Aqueous Media

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Received: 27 February 2013 / Accepted: 30 April 2013 / Published online: 8 May 2013 © Springer Science+Business Media New York 2013

Abstract Two novel rhodamine derivatives were designed and synthesized. They were successfully characterized by HR-MS, ¹H NMR and ¹³C NMR. They were found to exhibit a reversible colorimetric response and exhibit high selectivity and sensitivity for Hg(II) ion over other commonly coexistent metal ions. Their selectivity is excellent, and the detection of Hg(II) at ppb level is possible. The colorimetric and fluorescent response to Hg(II) can be conveniently detected even by the naked eye, which provides a facile method for visual detection of Hg(II).

Keywords Rhodamine \cdot Hg²⁺ \cdot Fluorescence \cdot Off-on

Introduction

The design of artificial chemosensors for selective and sensitive quantification of biologically and environmentally

Electronic supplementary material The online version of this article (doi:10.1007/s10895-013-1233-7) contains supplementary material, which is available to authorized users.

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Department of Chemistry, Key Laboratory of Bioorganic Phosphorus Chemistry & Chemical Biology (Ministry of Education), Tsinghua University, Beijing 100084, China heavy- and transition-metal ions in solution has attracted wide-spread interests of chemists, biologists, clinical biochemists and environmental scientists [1-3]. Mercury is a highly toxic and widespread global pollutant [4]. The mercuric ion, Hg [II], combines with both inorganic and organic ligands, which can readily penetrate through biological membranes even at very low concentrations, the US Environmental Protection Agency (EPA) standard for the maximum allowable level of inorganic Hg^{2+} in drinking water is 2 ppb [5]. There have been reported several traditional analytical techniques for Hg²⁺ include atomic absorption spectroscopy, cold vapor atomic fluorescence spectrometry and gas chromatography. These methods, however, require not only complicated instrumentation but also a long measuring time. Therefore, the design and development of fluorescent probes for Hg²⁺ has attracted a great deal of attention [6-8].

Several fluorescent chemosensors for Hg²⁺ have been reported, in which their signals are transduced either through a Hg^{2+} -complexation [9–12] or through a Hg²⁺-induced desulfurization reaction [13–16]. The former mode has some shortcomings, such as emission quenching as signal output, cross-sensitivities with other metallic cations, poor biocompatibility and low watersolubility. Though the latter mode has high selectivity toward Hg²⁺, these chemosensors are irreversible and can result in another contamination (HgS). In addition, these irreversible chemosensors cannot be used to monitor both the increase and the decrease of Hg²⁺ concentration in the environment or some biological metabolite. Therefore, development of reversibly fluorescent chemosensors for Hg2+ determination with light "off-on" and high selectivity is still highly desirable.

On the basis of the well-known spirolactam (nonfluorescent) to open-ring amide(fluorescent) equilibrium, rhodamine frameworks have been considered an





ideal mode for the construction of the off-on systems that have frequently been utilized to design fluorescence-enhanced probes for metal ions [17–24]. Recently, our group has developed a series of rhodamine-based chemosensors to sense metal ions [25–27]. We have even reported a rhodamine-based chemosensor for Hg^{2+} by a desulfurization reaction [28]. However, the irreversible reaction restricts its biological application. In this paper, two rhodamine-based chemosensors were designed for reversible and highly selective Hg^{2+} determination with a light "off–on" switch (Scheme 1).

Experimental Section

Apparatus

Fluorescence spectra measurements were performed on a HITACHI F-4500 fluorescence spectrophotometer, and the excitation and emission wavelength band passes were both set at 4.0 nm. Absorption spectra were measured on a Lambda 35. UV/VIS spectrometer, Perkin Elmer precisely. The melting points were determined by an X-4 microscopic melting point apparatus with a digital thermometer (Shanghai, China). The pH was measured with a Model pHs-3C

Fig. 1 Absorbance spectra of 10 μ M L2 in the presence of 50 eq. different metal ions in CH₃OH/H₂O(9:1, v/v). Insert shows the photos of sensor L2 with different metal ions



Fig. 2 Job's plot evaluated from the absorption spectra of L2 and Hg²⁺ in CH₃OH/H₂O (9/1, v/v) solution. Inset: Job's plot of changes of absorbance at 558 nm, the total concentration of [Hg²⁺]+[L2] was 100 μ M



meter (Shanghai, China). ¹H and ¹³C NMR spectra were recorded using a Bruker DTX-400 spectrometer. Samples were dissolved in CDCl₃ and placed in 5 mm NMR tubes. TMS was used as internal reference. ESI mass spectra were carried out on an HPLC Q-T of HR-MS spectrometer (Waters Micromass) by using methanol as mobile phase.

Materials

All chemicals and reagents were used as received from commercial sources without further purification. Solvents for

Fig. 3 ESI mass spectra (positive) of **L2** in the presence of HgCl₂ (5 equiv), indicating the formation of a 1:1 metalligand complex chemical synthesis and analysis were purified according to standard procedures. Double distilled water was used throughout the experiment. Chloride salts of metal ions (K⁺, Na⁺, Ca²⁺, Mg²⁺, Ba²⁺, Zn²⁺, Fe³⁺, Mn²⁺, Pb²⁺, Cu²⁺, Co²⁺, Ni²⁺, Cd²⁺, Cr³⁺, Hg²⁺) and the nitrate salt of Ag⁺ ions were used to evaluate the metal ion binding properties by synthesized compounds. The metal ions were prepared as 10.00 mmol/L in water solution. The stoichiometry of each compound and Hg²⁺ was determined by Job's method from the obtained absorption spectroscopic data. In the determination, the sum of concentration of Hg²⁺ and each compound



was kept at 100 μM and the molar ratio of Hg^{2+} was changed from 0 to 1.0.

Experimental details can be found in the Supporting Information. Compounds 1 and 2 were conveniently synthesized according to the procedure as published in the literatures [25].

Synthesis of Compound 3

To a solution of bromoacetyl bromide (0.06 mL, 0.71 mmol) in dry dichloromethane(20 ml) was added $K_2CO_3(98mg, 0.71 mmol)$. The mixture was cooled in an ice bath and then compound **2**(235 mg, 0.47 mmol) dissolved in 15 ml dry dichloromethane was added dropwise over 30 min. The reaction mixture was stirred at ice bath for about 2 h. It



was washed successively with 2 % Na_2CO_3 water solution (3×30 mL). The organic layer was dried over Na_2SO_4 and evaporated under vacuo. The residue 3 obtained was directly used for next step without further purification.

Synthesis of Compound L2

To a stirred solution of compound 3(62 mg, 0.1 mmol)in dry toluene (30 mL), 1,4,7,10-macrocyclic polyamide (125 mg,0.725 mmol) was added. The solution was refluxed for about 48 h under dry N₂. The resulting solution was concentrated by evaporation and dissolved in 10 ml ethanol. Concentrated HCl was added to the solution, and the precipitated HCl salt of the unreacted cyclen was removed by filtration. The residue was



concentrated by evaporation, dissolved in 10 ml water, and washed with CHCl₃. Aqueous NaOH solution (5 M, 15 ml) was added to the solution and extracted with CH₂Cl₂ (10 ml×5). The organic layer was dried over Na₂SO₄ and concentrated by evaporation. The residue was dried in vacuo at 385 K for 8 h, affording L2 as a yellow powder. M.p: 92-94 °C. Yield of L2: 84 %. ¹H NMR (400 MHz, CDCl₃, ppm): 1.17 (t, 12 H, J= 6.8 Hz); 2.71 (m, 16 H); 3.06 (d, 4 H, J=12.8 Hz); 3.34 (q, 4 H, J=6.7 Hz); 3.68 (t, 2 H, J=7.2 Hz); 6.34 (m, 6 H); 7.09 (d, 1 H, J=4 Hz); 7.46 (s, 2 H); 8.16 (t, 1 H, J=18.6 Hz). ¹³C NMR (CDCl₃, 100 MHz, ppm); 12.61, 14.17, 22.71, 29.38, 31.94, 36.72, 40.04, 43.83, 44.38, 46.12, 46.66, 47.31, 47.91, 53.51, 59.55, 73.37, 73.59, 97.68, 97.99, 98.13, 103.32, 103.83, 108.15, 123.27, 124.89, 125.01, 128.41, 128.50, 128.70, 132.38, 132.52, 137.61,149.16, 151.49, 153.24, 171.62, 191.16. HR-MS: calcd for $[C_{40}H_{57}N_8O_2S^+]$: 713.4320. Found: 713.4324 [M+H]⁺.

Synthesis of Compound L1

Similar method, **L1** was obtained in 65 % yield. ¹H NMR (400 MHz, CDCl₃, ppm): 1.16 (t, 12 H, J=6.6 Hz); 2.95 (t, 2 H, J=7.8 Hz); 3.33 (q, 8 H, J=6.9 Hz); 3.78 (t, 2 H, J= 5.4 Hz); 4.50 (s, 2 H,); 6.20 (t, 4 H, J=9.4 Hz); 6.38(s, 2 H); 6.48 (s, 1 H); 7.04 (m, 3 H); 7.51 (s, 3 H); 8.15 (d, 1 H, J= 6.4 Hz); ¹³C NMR (CDCl₃, 100 MHz, ppm): 12.54, 29.68, 39.27; 42.78, 44.39, 50.10, 73.62, 97.79, 103.39, 108.51, 120.04, 123.28, 125.23, 128.19, 128.71, 129.81, 132.85, 137.62, 137.94, 149.28, 150.98, 153.13, 166.43, 193.06. ESI-MS: m/z 609 $[M+H]^+$; HR-MS: calcd for $[C_{35}H_{41}N_6O_2S^+]$: 609.3006. Found: 609.3007 $[M+H]^+$.

Fig. 5 The fluorescence

emission spectra of L2 (10 μ M) in the presence of different concentrations of Hg²⁺(0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 eq. respectively) in CH₃OH/H₂O (9/1, v/v), λ_{ex} =550 nm. *Inset*: Changes in the emission intensity at 580 nm

Results and Discussion

UV-vis Spectral Responses of L2

As shown in Fig. 1, UV–vis spectrum of **L2** (10 μ M) exhibited only very weak bands over 450 nm. Addition of 10 equiv Hg²⁺ into solution immediately resulted in a significant enhancement of absorbance at about 558 nm simultaneously the color change into red. Under the identical condition, no obvious response could be observed upon the addition of other ions including K⁺, Na⁺, Ca²⁺, Mg²⁺, Ba²⁺, Zn²⁺, Fe²⁺, Fe³⁺, Mn²⁺, Pb²⁺, Cu²⁺, Co²⁺, Ni²⁺, Cd²⁺, Cr³⁺(Fig. 1). The results demonstrated that **L2** was characteristic of high selectivity toward Hg²⁺ over other competitive metal ions.

To further investigate the interaction of Hg^{2+} and L2, an ultraviolet titration experiment was carried out (Fig. 2). To determine the stoichiometry of the mercury-ligand complex, Job's method for absorbance measurement was applied. As shown in Fig. 2 inset, the absorbance reached a maximum when the ratio of $[Hg^{2+}]/{[Hg^{2+}]+[L2]}$ was 0.5, indicating a 1:1 stoichiometry of the Hg^{2+} to L2 in the complex. To confirm the stoichiometry between L2 and Hg^{2+} ion, ESI-MS analysis was conducted. As shown in Fig. 3, Mass peaks at m/z 913.4 corresponding to $[L2+Hg^{2+}-H^+]^+$ (calcd= 913.4) and m/z 949.4 corresponding to $[L2+Hg^{2+}+C1^-]^+$ (calcd=949.3) were clearly observed when 5 equiv of Hg^{2+} was added to L2, which gave a remarkable evidence for the formation of the1:1 complex of Hg^{2+} with L2.

Fluorescence Spectral Responses of L2

As shown in Fig. 4a, L2 (10 μ M) showed a very weak fluorescence in the absence of metal ions. When 10 equiv.





Fig. 6 The fluorescence intensity at 579 nm of compound L2(0.1 μ M) as a function of the Hg²⁺ concentration (0–20 ppb) in CH₃OH/H₂O (9/ 1, v/v), λ_{ex} =550 nm

metal ions of Zn^{2+} , Mg^{2+} , Ca^{2+} , Cd^{2+} , Cu^{2+} , Pb^{2+} , Mn^{2+} , Fe^{3+} , Ba^{2+} , Ni^{2+} , Fe^{2+} , K^+ , Ag^+ , Co^{2+} , Cr^{3+} and Na^+ were added, no obvious changes on fluorescence intensity and color could be observed. However, under the same conditions, the addition of Hg^{2+} (10 equiv.) resulted in a remarkably enhancement of fluorescence (17-fold) at 579 nm. This strongly suggested that **L2** can serve as a high sensitivity for Hg^{2+} . Moreover, the enhancement in fluorescence intensity resulting from the addition of Hg^{2+} is not influenced significantly by the addition of the background metal ions (Fig. 4b).

To further investigate the binding stoichiometry of L2 and Hg^{2+} ion, a fluorescence titration experiment was carried out. An increase of fluorescence intensity of L2 could be observed with gradual addition of Hg^{2+} ion (Fig. 5). The increment saturated after adding 1.0 equiv of Hg^{2+} (Fig. 5, inset), this also indicating a 1:1 stoichiometry of the Hg^{2+} to L2 in the complex.

The Detection of Hg²⁺

Generally, one of the most important and useful applications for a fluorescent chemosensor is the detection of metal ions, especially for heavy metals. The limit of the chemosensor for Hg^{2+} ion has been tested. When L2 was employed at 0.1 μ M in CH₃OH/H₂O (9/1, v/v, pH 7.0), the fluorescent intensity of L2 was proportional to the concentration of Hg^{2+} added (Fig. 6). The detection limit was measured to be 1.0 ppb, establishing that L2 was capable of distinguishing safe and toxic levels of Hg^{2+} in drinking water according to the U.S. (EPA) standard (2 ppb).

Mechanism

Cyclen (1,4,7,10-tetraazacyclododecane) is good hydrophilic groups and can coordinate strongly with many transition metal cations. In order to improve target compound's water solubility, we introduce 1,4,7,10-macrocyclic polyamide into rhodamine. It is well-known that cyclen coordinates with one metal cation in organic media. Many metal cation can coordinate with cyclen, but the binding constant is low [29–31]. In aqueous media, coordination of cyclen nitrogens with the Hg²⁺ may weaker than that with water. As compound L2 contained a macrocycle and an "S" group, based on the coordination of Hg²⁺ to the S atom, we assumed that L2 may display a more fluorescence enhancement.

Considering the behaviors of absorption and fluorescence spectra, the turn-on response of L2 may be explained by the spirocycle open-close mechanism. The free probe L2 is the spirocyclic form, which is colorless and non fluorescent, whereas the coordination of Hg²⁺ leads to the spirocycle opening, resulting in an appearance of visible absorption and fluorescence. According to our knowledge [24], we propose a plausible mechanism which is illustrated in Scheme 2. In order to certify this binding model, we designed and synthesized compound L1 which contained an imidazole group, and the colorimetric and fluorescent results of L1 confirmed this binding model. The ultraviolet titration experiment, fluorescence titration experiment and ESI-MS all indicated a 1:1 stoichiometry of the Hg^{2+} to L1 or L2. That means the imidazole or cyclen group has no coordination with Hg^{2+} ion.

The little discrepancy in recognizing Hg^{2+} of L1 and L2 may duo to the different solubility of cyclen (1,4,7,10-tetraazacyclododecane) and imidazole in water. As the macrocycle had a better solubility than imidazole in water,

Scheme 2 Proposed mechanism for the fluorescence enhancement of L2 upon the addition of Hg^{2+}



spirolactam form

ring-open amide form

and this may display a beneficial effect to the sensing effect of L2 with Hg^{2+} in aqueous media.

Another direct evidence was obtained by comparing the IR spectra of L2 and L2-Hg(II) in KBr disks, and the results were shown in Figs.S13 and S14. The peak at 1,671 cm⁻¹, which corresponds to the amide carbonyl(C=O) absorption drastically shifted to lower frequency upon the addition of excess Hg(II), indicating that the carbonyl oxygen coordinated with Hg²⁺.

Further, it was of great interest to investigate the reversible binding nature of the sensor. To demonstrate the reversibility of L2, KI (10 eq.), as a strong affinity for Hg²⁺, was introduced into the solution containing L2(10 μ M) and Hg²⁺(100 μ M). Upon the addition of KI, the color of the mixture of L2 and Hg²⁺ changed from red to colorless and the fluorescence intensity was decreased, indicating that Γ sequestered Hg²⁺ of the mixture, liberated the free L2(Fig.S12). Thus, L2 can be classified as a reversible chemosensor not a chemodosimeter for Hg²⁺ ions.

Conclusion

In conclusion, we have developed two efficient rhodamine thiospirolactam-based fluorescent reversible chemosensor for Hg^{2+} . They exhibited a strong fluorescence enhancement upon addition of Hg^{2+} while showing almost no response to other cations. The significant changes in the fluorescence color could be used for naked-eye detection, the limit of detection of **L2** for Hg^{2+} in CH₃OH/H₂O (9/1, v/v) was found to be 1 ppb. They may therefore be applicable as rhodamine-based turn-on type fluorescent chemosensors for Hg^{2+} .

Acknowledgments This work was financially supported by the National Science Foundation of China (Nos. 20972143 and 20972130) and Program for New Century Excellent Talents in University (NCET-11-0950).

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